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**Priority Certificate
for the filing of a Patent Application**

File Reference: 198 19 829.9

Filing date: 4 May 1998

Applicant/Proprietor: Bayer Aktiengesellschaft, Leverkusen/DE

Title: Nucleic acids which encode insect acetylcholine receptor subunits

IPC: C 07 K, A 61 K, C 12 N

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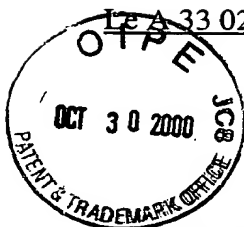
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- 1 -

Nucleic acids which encode insect acetylcholine receptor subunits

5 The invention relates, in particular, to nucleic acids which encode insect acetylcholine receptor subunits.

10 Nicotinic acetylcholine receptors are ligand-regulated ion channels which are of importance in neurotransmission in the animal kingdom. The binding of acetylcholine or other agonists to the receptor induces a transient opening of the channel and allows cations to flow through. It is assumed that a receptor consists of five subunits which are grouped around a pore. Each of these subunits is a protein which consists of an extracellular N-terminal moiety followed by three transmembrane regions, an intracellular moiety, a fourth transmembrane region and a short extracellular C-terminal moiety (Changeux et al. 1992).

15 Acetylcholine receptors are especially well investigated in vertebrates. In this context, three groups can be distinguished on the basis of their anatomical location and their functional properties (conducting properties of the channel, desensitization, and sensitivity towards agonists and antagonists and also towards toxins such as α -bungarotoxin). The classification correlates with the molecular composition of the receptors. There are heterooligomeric receptors having the subunit composition $\alpha_2\beta\gamma\delta$, which are found in muscle (Noda et al. 1982, Claudio et al. 1983, Devillers-Thiery et al. 1983, Noda et al. 1983a, b), heterooligomeric receptors which contain subunits from the $\alpha_2 - \alpha_6$ and $\beta_2 - \beta_4$ groups and which are found in the nervous system (Wada et al. 1988, Schoepfer et al. 1990, Cockcroft et al. 1991, Heinemann et al. 1997), and also homooligomeric receptors which contain subunits from the $\alpha_7 - \alpha_9$ group and which are likewise found in the nervous system (Lindstrom et al. 1997, Elgoyhen et al. 1997). This classification is also supported by an examination of the relatedness of the gene sequences of the different subunits. Typically, the sequences of functionally homologous subunits from different species are more similar to each other than are sequences of subunits which are from different groups but from the same species. Thus, the rat muscle α subunit, for example, exhibits 78% amino acid identity and 84% amino acid similarity with that of the electric ray *Torpedo californica* but only 48% identity and 59% similarity with the rat α_2 subunit (heterooligomeric, neuronal) and 36% identity and 45% similarity with the rat α_7 subunit (homooligomeric, neuronal). Furthermore, the gene sequences of all the

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known acetylcholine receptor subunits are to a certain extent similar not only to each other but also to those of some other ligand-regulated ion channels (e.g. the serotonin receptors of the 5HT₃ type, the GABA-regulated chloride channels and the glycine-regulated chloride channels). It is therefore assumed that all these receptors are descended from one common precursor and they are classified into one supergene family (Ortells et al. 1995).

In insects, acetylcholine is the most important excitatory neurotransmitter of the central nervous system. Accordingly, acetylcholine receptors can be detected electrophysiologically in preparations of insect central nervous system ganglia. The receptors are detected both in postsynaptic and presynaptic nerve endings and in the cell bodies of interneurons, motor neurons and modulatory neurons (Breer et al. 1987, Buckingham et al. 1997). Some of the receptors are inhibited by α -bungarotoxin while others are insensitive (Schloß et al. 1988). In addition, the acetylcholine receptors are the molecular point of attack for important natural (e.g. nicotine) and synthetic insecticides (e.g. chloronicotinyls).

The gene sequences of a number of insect nicotinic acetylcholine receptors are already known. Thus, the sequences of five different subunits have been described in *Drosophila melanogaster* (Bossy et al. 1988, Hermanns-Borgmeyer et al. 1986, Sawruk et al. 1990a, 1990b, Schulz et al. Unpublished, EMBL accession number Y15593), while five have likewise been described in *Locusta migratoria* (Stetzer et al. unpublished, EMBL accession numbers AJ000390 - AJ000393), one has been described in *Schistocerca gregaria* (Marshall et al. 1990), two have been described in *Myzus persicae* (Sgard et al. unpublished, EMBL accession number X81887 and X81888), and one has been described in *Manduca sexta* (Eastham et al. 1997). Furthermore, a number of partial gene sequences from *Drosophila melanogaster* have been characterized as so-called expressed sequence tags (Genbank accession numbers AA540687, AA698155, AA697710, AA697326). The fact that individual sequences are very similar to those from other insects suggests that these subunits are functional homologues.

It is of great practical importance to make available new insect acetylcholine receptor subunits, for example for the purpose of searching for novel insecticides, with those subunits which differ from the known subunits to a greater extent than is the case between functional homologues being particularly of interest.

The present invention is consequently based, in particular, on the object of making available nucleic acids which encode novel insect acetylcholine receptor subunits.

5 This object is achieved by the provision of nucleic acids which comprise a sequence selected from

- 10 (a) the sequences according to SEQ ID NO: 1, SEQ ID NO: 3 or SEQ ID NO: 5,
- (b) part sequences of the sequences defined in (a) which are least 14 base pairs in length,
- 15 (c) sequences which hybridize to the sequences defined in (a) in 2 x SSC at 60°C, preferably in 0.5 x SSC at 60°C, particularly preferably in 0.2 x SSC at 60°C (Sambrook et al. 1989),
- 20 (d) sequences which exhibit at least 70% identity with the sequences defined in (a), between position 1295 and position 2195 in the case of SEQ ID NO: 1, or between position 432 and position 1318 in the case of SEQ ID NO: 3, or between position 154 and position 1123 in the case of SEQ ID NO: 5,
- 25 (e) sequences which are complementary to the sequences defined in (a), and
- (f) sequences which, because of the degeneracy of the genetic code, encode the same amino acid sequences as the sequences defined in (a) to (d).

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The degree of identity of the nucleic acid sequences is preferably determined using the GAP program from the GCG program package, Version 9.1 with standard settings.

35 The present invention is based on the surprising finding that insects possess genes which encode subunits of, in particular, homooligomeric acetylcholine receptors.

The invention furthermore relates to vectors which contain at least one of the novel nucleic acids. All the plasmids, phasmids, cosmids, YACs or artificial chromosomes which are used in molecular biological laboratories can be used as vectors. These vectors can be linked to the usual regulatory sequences for the purpose of expressing the novel nucleic acids. The choice of such regulatory sequences depends on whether prokaryotic or eukaryotic cells, or cell-free systems, are used for the expression. The SV40, adenovirus or cytomegalovirus early or late promoter, the lac system, the trp system, the main operator and promoter regions of phage lambda, the control regions of the fd coat protein, the 3-phosphoglycerate kinase promoter, the acid phosphatase promoter and the yeast α -mating factor promoter are examples of expression control sequences which are particularly preferred.

In order to be expressed, the nucleic acids according to the invention can be introduced into suitable host cells. Both prokaryotic cells, preferably E.coli, and eukaryotic cells, preferably mammalian or insect cells, are suitable for use as host cells. Other examples of suitable unicellular host cells are: Pseudomonas, Bacillus, Streptomyces, yeasts, HEK-293, Schneider S2, CHO, COS1 and COS7 cells, plant cells in cell culture and also amphibian cells, in particular oocytes.

The present invention also relates to polypeptides which are encoded by the nucleic acids according to the invention and also the acetylcholine receptors, preferably homooligomeric acetylcholine receptors, which are synthesized from them.

In order to prepare the polypeptides which are encoded by the nucleic acids according to the invention, host cells which contain at least one of the nucleic acids according to the invention can be cultured under suitable conditions. After that, the desired polypeptides can be isolated from the cells or the culture medium in a customary manner.

The invention furthermore relates to antibodies which bind specifically to the abovementioned polypeptides or receptors. These antibodies are prepared in the customary manner. For example, such antibodies can be produced by injecting a substantially immunocompetent host with a quantity of an acetylcholine receptor polypeptide, or a fragment thereof, according to the invention which is effective for producing antibodies, and subsequently isolating these antibodies. Furthermore, an immortalized cell line which produces monoclonal antibodies can be obtained in a

manner known per se. Where appropriate, the antibodies can be labelled with a detection reagent. Preferred examples of such a detection reagent are enzymes, radioactively labelled elements, fluorescent chemicals or biotin. Instead of the complete antibody, use can also be made of fragments which possess the desired specific binding properties.

The nucleic acids according to the invention can be used, in particular, for producing transgenic invertebrates. These latter can be employed in test systems which are based on an expression of the receptors according to the invention, or variants thereof, which differs from that of the wild type. In addition, this includes all transgenic invertebrates in which a change in the expression of the receptors according to the invention, or their variants, occurs as the result of modifying other genes or gene control sequences (promoters).

The transgenic invertebrates are produced, for example, in *Drosophila melanogaster* by means of P element-mediated gene transfer (Hay et al., 1997) or in *Caenorhabditis elegans* by means of transposon-mediated gene transfer (e.g. using Tc1, Plasterk, 1996).

The invention also consequently relates to transgenic invertebrates which contain at least one of the nucleic acid sequences according to the invention, preferably to transgenic invertebrates of the species *Drosophila melanogaster* or *Caenorhabditis elegans*, and to their transgenic progeny. Preferably, the transgenic invertebrates contain the receptors according to the invention in a form which differs from that of the wild type.

The nucleic acids according to the invention can be prepared in the customary manner. For example, the nucleic acid molecules can be synthesized entirely chemically. In addition, only short segments of the sequences according to the invention can be synthesized chemically and these oligonucleotides can be labelled radioactively or with a fluorescent dye. The labelled oligonucleotides can be used to screen cDNA libraries prepared from insect mRNA. Clones which hybridize to the labelled oligonucleotides are selected for isolating the relevant DNA. After the isolated DNA has been characterized, the nucleic acids according to the invention are readily obtained.

The nucleic acids according to the invention can also be prepared by means of PCR methods using chemically synthesized oligonucleotides.

The nucleic acids according to the invention can be used for isolating and characterizing the regulatory regions which occur naturally adjacent to the coding region. Consequently, the present invention also relates to these regulatory regions.

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The nucleic acids according to the invention can be used to identify novel active compounds for plant protection, such as compounds which, as modulators, in particular as agonists or antagonists, alter the conducting properties of the acetylcholine receptors according to the invention. For this, a recombinant DNA molecule, which encompasses at least one nucleic acid according to the invention, is introduced into a suitable host cell. The host cell is cultured, in the presence of a compound or a sample which comprises a multiplicity of compounds, under conditions which permit expression of the receptors according to the invention. A change in the receptor properties can be detected, as described below in Example 2. Using this approach, it is possible to discover insecticidal substances.

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The nucleic acids according to the invention also make it possible to discover compounds which bind to the receptors according to the invention. These compounds can likewise be used as insecticides on plants. For example, host cells which contain the nucleic acid sequences according to the invention and express the corresponding receptors or polypeptides, or the gene products themselves, are brought into contact with a compound or a mixture of compounds under conditions which permit the interaction of at least one compound with the host cells, receptors or the individual polypeptides.

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Host cells or transgenic invertebrates which contain the nucleic acids according to the invention can also be used to discover substances which alter the expression of the receptors.

The above-described nucleic acids, vectors and regulatory regions according to the invention can additionally be used for discovering genes which encode polypeptides which are involved in the synthesis, in insects, of functionally similar acetylcholine receptors. According to the present invention, functionally similar receptors are understood as being receptors which encompass polypeptides which, while differing in their amino acid sequences from the polypeptides described in this present publication, essentially possess the same functions.

35

Comments on the sequence listing and the figures:

5 SEQ ID NO: 1 shows the nucleotide sequence of the isolated Da7 cDNA, beginning with position 1 and ending with position 2886. SEQ ID NO: 1 and SEQ ID NO: 2 also show the amino acid sequences of the protein deduced from the Da7 cDNA sequence.

10 SEQ ID NO: 3 shows the nucleotide sequence of the isolated Hva7-1 cDNA, beginning with position 1 and ending with position 3700. SEQ ID NO: 3 and SEQ ID NO: 4 also show the amino acid sequences of the protein deduced from the Hva7-1 cDNA sequence.

15 SEQ ID NO: 5 shows the nucleotide sequence of the isolated Hva7-2 cDNA, beginning with position 1 and ending with position 3109. SEQ ID NO: 5 and SEQ ID NO: 6 also show the amino acid sequences of the protein deduced from the Hva7-2 cDNA sequence.

20 Figure 1 shows the increase in intracellular calcium which occurs in cells which have been recombinantly modified as described in Example 2 following the addition of nicotine. Cells were loaded with Fura-2-acetoxymethyl ester (5 - 10 μ M in serum-free minimal essential medium containing 1% bovine serum albumin and 5 mM calcium chloride), washed with Tyrode solution buffered with N-(2-hydroxyethyl)piperazine-N'-(2-ethanesulphonic acid) (5 mM HEPES) and alternately illuminated, under a fluorescence microscope (Nikon Diaphot) with light of 340 nm and 380 nm wavelength. A measurement point corresponds to a pair of video images at the two wavelengths (exposure time per image, 100 ms). The time interval between two measurement points is 3 s. After 8 images had been taken (measurement point 4.0), nicotine was added to a final concentration of 500 μ M and the measurement series was continued. The fluorescence intensity of the cells when
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30 illuminated with light of 380 nm wavelength was divided by the corresponding intensity at 340 nm, thereby giving the ratio.

Examples:

Example 1

- 5 Isolating the described polynucleotide sequences

Polynucleotides were manipulated using standard methods of recombinant DNA technology (Sambrook, et al., 1989). The bioinformatic processing of nucleotide and protein sequences was carried out using the GCG program package Version 9.1
10 (GCG Genetics Computer Group, Inc., Madison Wisconsin, USA).

Partial polynucleotide sequences

Sequence comparisons ("Clustalw") were used to identify regions, from which
15 degenerate oligonucleotides were deduced by backtranslating the codons, of protein sequences from genes whose ability to form homooligomeric acetylcholine receptors was known. In all, 5 such oligonucleotide pairs were selected for the polymerase chain reaction (PCR). Only one combination (see below) gave a product both from Heliothis cDNA and from Drosophila cDNA.

20 RNA was isolated from whole Heliothis virescens embryos (shortly before hatching) using Trizol reagent (Gibco BRL, in accordance with the manufacturer's instructions). The same procedure was adopted with Drosophila embryos (24 h at 25°C). 10 µg of these RNAs were employed in a first cDNA strand synthesis
25 (Superscript Preamplification System for first cDNA strand synthesis, Gibco BRL, in accordance with the manufacturer's instructions, reaction temperature 45°C).

Subsequently, 1/100 of the abovementioned first-strand cDNA was in each case employed in a polymerase chain reaction (PCR) using the oligonucleotides alpha7-1s: (5'-GAYGTIGAYGARAARAAYCA-3') and alpha7-2a: (5'-
30 CYYTCRTCIGCRCTRTRTA-3') (recombinant Taq DNA polymerase, Gibco BRL). The PCR parameters were as follows: Hva7-1 and Hva7-2: 94°C, 2 min; 35 times (94°C, 45 s; 50°C, 30 s; 72°C, 60 s) and also Da7: 96°C, 2 min; 35 times (96°C, 45 s; 50°C, 30 s; 72°C, 60 s). In each case, this resulted in a dectable band of
35 approx. 0.2 kb in an agarose gel (1%), both in the case of Drosophila cDNA and in the case of Heliothis cDNA. After the DNA fragments had been subcloned by means

of SrfScript (Stratagene), and their sequences had been determined, it turned out that two different DNA fragments had been amplified from *Heliothis* cDNA; these were 228-11 = Hva7-1 (partial, containing 165 bp) and 228-8 = Hva7-2 (partial, containing 171 bp). Only one DNA fragment was isolated from *Drosophila* cDNA; this was 248-5 = Da7 (partial, containing 150 bp).

Isolating poly A-containing RNA from *Heliothis virescens* tissue and constructing the cDNA libraries

The RNA for cDNA library I was isolated from whole *Heliothis virescens* embryos (shortly before hatching) using Trizol reagent (Gibco BRL, in accordance with the manufacturer's instructions). The RNA for cDNA library II was isolated from whole head ganglia from 500 *Heliothis virescens* larvae (stages 4-5) using Trizol reagent (Gibco BRL, in accordance with the manufacturer's instructions). The poly A-containing RNAs were then isolated from these RNAs by purifying with Dyna Beads 280 (Dyna). 5 µg of these poly A-containing RNAs were subsequently employed in constructing cDNA libraries I and II using the λ-ZAPExpress vector (cDNA Synthesis Kit, ZAP-cDNA Synthesis Kit and ZAP-cDNA Gigapack III Gold Cloning Kit, all from Stratagene). In a departure from the manufacturer's instructions, Superscript Reverse Transcriptase (Gibco BRL) was used for synthesizing the cDNA at a synthesis temperature of 45°C. In addition, radioactively labelled deoxynucleoside triphosphates were not added. Furthermore, the synthesized cDNAs were not fractionated through the gel filtration medium contained in the kit but instead through Size Sep 400 Spun Columns (Pharmacia).

Complete polynucleotide sequences

Apart from the first screening round when isolating the Hva7-1 clone, all the screens were carried out using the DIG system (all reagents and consumables from Boehringer Mannheim, in accordance with the instructions in "The DIG System User's Guide for Filter Hybridization", Boehringer Mannheim). The DNA probes employed were prepared by means of PCR using digoxigenin-labelled dUTP. The hybridizations were carried out at 42°C overnight in DIG Easy Hyb (Boehringer Mannheim). Labelled DNA was detected on nylon membranes by means of chemiluminescence (CDP-Star, Boehringer Mannheim) using X-ray films (Hyperfilm MP, Amersham). Initial partial sequencing of the isolated gene library plasmids was

carried out, for identification purposes, using T3 and T7 primers (ABI Prism Dye Terminator Cycle Sequencing Kit, ABI, using an ABI Prism 310 Genetic Analyzer). The complete polynucleotide sequences in Hva7-1, Hva7-2 and Da7 were determined, as a commissioned sequencing carried out by Qiagen, Hilden, by means of primer walking using cycle sequencing.

a. Isolating the Da7 clone

10^6 phages from a *Drosophila melanogaster* cDNA library in λ phages (Canton-S embryo, 2-14 hours, in Uni-ZAP XR vector, Stratagene) were screened using DIG-labelled 248-5 as the probe (in accordance with the manufacturer's (Stratagene) instructions). The maximum stringency when washing the filters was: 0.2 x SSC; 0.1% SDS; 42°C; 2 x 15 min. One clone (clone 432-1) was isolated whose insert had a size of 2940 bp (Da7, SEQ ID NO: 1). The largest open reading frame of this sequence begins at position 372 of the depicted sequence and ends at position 1822. The 770 amino acid polypeptide sequence which is deduced from this (SEQ ID NO: 2) encodes a protein having a calculated molecular weight of 87.01 kD.

b. Isolating the Hva7-1 clone

10^6 phages from the *Heliothis virescens* embryo cDNA library (library I) were included in the screening. The first of three screening rounds took place using α - ^{32}P -labelled 228-11 DNA as the probe. The probe was hybridized to the filters in Quickhyb (Stratagene) at 68°C for one hour. The filters were then washed twice, for 15 min on each occasion, at room temperature in 2 x SSC; 0.1% SDS and twice, for 30 min on each occasion, at 42°C in 0.1xSSC; 0.1% SDS. Hybridized probe was detected by means of autoradiography, at -80°C overnight, using XR X-ray films (Kodak) and employing intensifying screens (Amersham). The two further screening rounds were carried out using the DIG System (Boehringer Mannheim).

The clone 241-5, which was isolated in this screen, contained an insert of 3630 bp. This insert (Hva7-1, SEQ ID NO: 3) possesses a longest open reading frame which begins at position 335 of the depicted nucleic acid sequence and ends at position 1821. The 496 amino acid polypeptide which is deduced from this (SEQ ID NO: 4) encodes a protein having a calculated molecular weight of 56.36 kD.

c. Isolating the Hva7-2 clone

10⁶ phages from the *Heliothis virescens* ganglia cDNA library (library II) were included in the screening. Dig-labelled 228-8 DNA was used as the probe. The maximum stringency when washing the filters was: 0.1 x SSC; 0.1% SDS; 42°C; 2 x 15 min.

The clone 241-5, which was isolated in this screen, contained an insert of 3630 bp. This insert (Hva7-2, SEQ ID NO: 5) possesses a longest open reading frame which begins at position 95 of the depicted nucleic acid sequence and ends at position 1598. The 501 amino acids polypeptide which was deduced from this (SEQ ID NO: 6) encodes a protein having a calculated molecular weight of 56.71 kD.

Example 2

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Generating the expression constructs

a. Da7

20 The sequence region from position 372 to position 2681 of SEQ ID NO: 1 was amplified by means of a polymerase chain reaction (PCR). Deoxyoligonucleotides having the sequences

GCGAATTCACCACCATGAAAAATGCACAACCTG

and

CGAGACAATAATATGTGGTGCCTCGAG were used for this. The Pfu

25

polymerase from Stratagene was used as the DNA polymerase in accordance with the manufacturer's instructions.

Following the amplification, the segment which had been generated was digested with the restriction endonucleases Eco RI and Xho I and cloned into a vector, i.e. pcDNA3.1/Zeo (Invitrogen), which had likewise been digested with Eco RI and Xho I.

30

b. Hva7-1

The sequence region from position 335 to position 1822 from SEQ ID NO: 3 was amplified by means of a polymerase chain reaction (PCR). Deoxyoligonucleotides having the sequences

35

GCAAGCTTACCACCATGGGAGGTAGAGCTAGACGCTCGCAC and
GCCTCGAGCGACACCATGATGTGTGGCGC were used for this. The Pfu
polymerase from Stratagene was used as the DNA polymerase in accordance with the
manufacturer's instructions.

5

Following amplification, the generated segment was digested with the restriction
endonucleases HindIII and Xho I and cloned into a vector, i.e. pcDNA3.1/Zeo
(Invitrogen), which had likewise been digested with HindIII and Xho I.

10 c. Hva7-2

The sequence region from position 95 to position 1597 from SEQ ID NO: 5 was
amplified by means of a polymerase chain reaction (PCR). Deoxyoligonucleotides
having the sequences

15 GCAAGCGCCGCTATGGCCCCTATGTTG and
TTGCACGATGATATGCGGTGCCTCGAGCG were used for this. The Pfu
polymerase from Stratagene was used as the DNA polymerase in accordance with the
manufacturer's instructions. Following amplification, the generated segment was
digested with the restriction endonucleases HindIII and Xho I and cloned into a
20 vector, i.e. pcDNA3.1/Zeo (Invitrogen), which had likewise been digested with
HindIII and Xho I.

d.Hva7-1 / 5HT₃ and Hva7-2 / 5HT₃ chimaeras

25 The region from position 335 to position 1036 from SEQ ID NO: 3 (Hva7-1/5HT₃
chimaera) and the region from position 95 to position 763 from SEQ ID NO: 5
(Hva7-2/5HT₃ chimaera) was in each case fused to the region from position 778 to
position 1521 from the Mus musculus 5-HT₃ receptor cDNA (sequence in EMBL
database: M774425) using the method of overlap extension (Jespersen et al. 1997).
30 The two fragments were subsequently cloned into the pcDNA3.1/Zeo vector by
means of TA cloning (Invitrogen, in accordance with the manufacturer's
instructions). Constructs containing the correct orientation of the two fragments in
the vector were identified by sequencing using the T7 primer (Invitrogen).

Cell culture and gene transfer

HEK293 cells, which express the α subunit of an L-type Ca channel (Zong et al. 1995, Stetzer et al. 1996), were cultured in Dulbecco's modified Eagle's medium and 10% foetal calf serum at 5% CO₂ and from 20°C to 37°C. FuGENE 6 (Boehringer Mannheim GmbH, Mannheim, Germany) was used for the gene transfer in accordance with the manufacturer's instructions. At from 24 h to 48 h after the gene transfer, the cells were sown at various densities in microtitre plates. Recombinantly altered cells were selected by growth in Dulbecco's modified Eagle's medium and 10% foetal calf serum and 150 - 500 µg/ml of Zeocin/ml over a period of from 3 to 4 weeks. Individual resistant clones were analyzed as described below.

Fura-2 measurements

The alterations in the intracellular calcium concentration were measured using Fura-2. A stock solution containing 2 mM Fura-2-acetoxy methyl ester (Sigma) in dimethyl sulphoxide (DMSO) was diluted to a final concentration of 5 - 10 µM in serum-free minimal essential medium (MEM, Gibco) containing 1% bovine serum albumin and 5 mM calcium chloride. The cells were incubated for from 45 to 60 min in this solution in a microtitre plate. The cells were then washed twice in Tyrode solution buffered with N-(2-hydroxyethyl)piperazine-N'-(2-ethanesulphonic acid) (5 mM HEPES) (HEPES-buffered salt solution containing 130 mM NaCl, 5 mM KCl, 2 mM CaCl₂, 1 mM MgCl₂, 5 mM NaHCO₃, 10 mM glucose). 100 µl Tyrode buffer were added to the wells of the microtitre plate and the cells were illuminated alternately, under a fluorescence microscope (Nikon Diaphot), with light of 340 nm and 380 nm wavelength. A series of video images (exposure time per image 100 ms) were taken with pauses of 3 seconds and stored, as digitalized images, in an image analysis computer (Leica, Quantimet 570). After 8 images had been taken (measurement point 4.0 in Fig. 1), nicotine was added to a final concentration of 500 µM and the measurement series was continued. The fluorescence intensity of the cells when illuminating with light of 380 nm wavelength was divided by the corresponding intensity at 340 nm and in this way a ratio was formed which represents the relative increase in calcium concentration (Grynkiewicz et al. 1985).

References:

- 5 Bossy et al. (1988) Conservation of neural nicotinic acetylcholine receptors from
Drosophila to vertebrate central nervous systems, EMBO J. 7, 611-618
- Breer et al. (1987) Molecular properties and functions of insect acetylcholine
receptors, J. Insect Physiol. 33, 771-790
- 10 Buckingham et al. (1997) Imidacloprid actions on insect neuronal acetylcholine
receptors, J.Exp. Biol. 200, 2685-2692
- Changeux et al. (1992) The functional architecture of the nicotinic acetylcholine
receptor explored by affinity labelling and site-directed mutagenesis, Quarterly
Review of Biophysics 25, 395-432
- 15 Claudio et al. (1983) Nucleotide and deduced amino acid sequences of Torpedo
californica acetylcholine receptor α subunit, Proc. Natl. Acad. Sci. USA 80, 1111-
1115
- 20 Devillers-Thiery et al. (1983) Complete mRNA coding sequence of the acetylcholine
binding α -subunit of Torpedo marmorata acetylcholine receptor: a model for the
transmembrane organization of the polypeptide chain, Proc. Natl. Acad. Sci. USA 80,
2067-2071
- 25 Elgoyhen et al. (1997) US Pat. No. 5,683,912
- Eastham et al. (1998) Characterisation of a nicotinic acetylcholine receptor from the
insect Manduca sexta, Eur. J. Neurosci 10, 879-889
- 30 Grynkiewicz et al. (1985) A new generation of Ca^{2+} indicators with greatly
improved fluorescence properties, J. Biol. Chem. 260, 3440-3450
- Hay et al. (1997), P element insertion-dependent gene activation in the Drosophila eye,
Proceedings of The National Academy of Sciences of The United States of America 94
35 (10), 5195-5200

Hermans-Borgmeyer et al. (1986) Primary structure of a developmentally regulated nicotinic acetylcholine receptor protein from *Drosophila* EMBO J. 5, 1503-1508

Heinemann et al. (1997) US Pat. No 5,591,590

5

Jespersen et al. (1997) Efficient Non-PCR-Mediated Overlap Extension of PCR Fragments by Exonuclease "End Polishing", *Biotechniques*, 23, 48-52

Lindstrom et al. (1997) US Pat. No. 5,599,709

10

Marshall et al. (1990) Sequence and functional expression of a single α subunit of an insect nicotinic acetylcholine receptor, EMBO J. 9, 4391-4398

Noda et al. (1982), Primary structure of α -subunit precursor of *Torpedo californica* acetylcholine receptor deduced from cDNA sequence, *Nature* 299, 793-797

15

Noda et al. (1983a), Primary structures of β - and δ -subunit precursor of *Torpedo californica* acetylcholine receptor deduced from cDNA sequences, *Nature* 301, 251-255

20

Noda et al. (1983b), Structural homology of *Torpedo californica* acetylcholine receptor subunits, *Nature* 302, 528-532

Ortells et al. (1995), Evolutionary history of the ligand-gated ion-channel superfamily of receptors, *Trends in Neurosciences* 18, 121-127

25

Plasterk (1996), The Tc1/mariner transposon family, *Transposable Elements, Current Topics in Microbiology and Immunology* 204, 125-143

Sambrook et al. (1989), *Molecular Cloning, A Laboratory Manual*, 2nd ed. Cold Spring Harbour Press

30

Sawruk et al. (1990a), EMBO J. 9, 2671-2677 Heterogeneity of *Drosophila* nicotinic acetylcholine receptors: SAD, a novel developmentally regulated α -subunit

35

Sawruk et al. (1990b), SBD, a novel structural subunit of the *Drosophila* nicotinic acetylcholine receptor, shares its genomic localization with two α -subunits, FEBS Lett. 273, 177-181

5 Schloß et al. (1988), Neuronal acetylcholine receptors of *Drosophila*: the ARD protein is a component of a high-affinity α -bungarotoxin binding complex, EMBO J. 7, 2889-2984

10 Stetzer et al. (1996) Stable expression in HEK-293 cells of the rat $\alpha 3/\beta 4$ subtype of neuronal nicotinic acetylcholine receptor, FEBS Lett. 397, 39-44

Zong et al. (1995) On the regulation of the expressed L-type calcium channel by cAMP-dependent phosphorylation, Pflügers Arch. - Eur. J. Physiol. 430, 340-347.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT:

- (A) NAME: Bayer Aktiengesellschaft
- (B) STREET: Bayerwerk
- (C) CITY: Leverkusen
- (E) COUNTRY: Germany
- (F) POSTAL CODE: D 51368

(ii) TITLE OF THE INVENTION: Nucleic acids which encode insect acetylcholine receptor subunits

(iii) NUMBER OF SEQUENCES: 6

(iv) COMPUTER-READABLE FORM:

- (A) MEDIUM TYPE: floppy disk
- (B) COMPUTER: IBM PC compatible
- (C) OPERATING SYSTEM: PC-DOS/MS-DOS
- (D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPA)

(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2886 base pairs
- (B) TYPE: nucleotide
- (C) STRANDEDNESS: double strand
- (D) TOPOLOGY: linear

(ii) TYPE OF MOLECULE: cDNA of mRNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Drosophila melanogaster

(vii) IMMEDIATE SOURCE:

- (B) CLONE(S): Da7

(ix) FEATURES:

- (A) NAME/KEY: CDS
- (B) POSITION: 372..2681

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

GGCACGAGAA AAAGTTGTGG TATAAACTTT TATTGTAGGA AAACGCATAA AAATAATAGA	60
AAAACGCTCT TCGGGTTGTA AAGAAAATAA GAAGACAAAA GAAAGACATG AAAACGTTGC	120
AAACAATAAA GCATATACTT GCCATATTGA TATAAAGGGA AATCGTGAAA AGGCGGTGAA	180
AATTCGTAA GATTAGTTGG TATTAAGGGC AGCCCATGCA CACAGCTAAA AAGGGA ACTA	240

AAAAAACCCC GCACAGAACA ATGAAAGCTG CAGCAGCTGG ATAAGGCCGA CAAAACCGAA	300
AATTATATTA TTGTAATCTA GTAGAGAGCA GACAACATAT CCGCTGGCAA CAACCAACAC	360
CGAAAGAGAC T ATG AAA AAT GCA CAA CTG AAA CTG ACT GAA GTT GAC GAT Met Lys Asn Ala Gln Leu Lys Leu Thr Glu Val Asp Asp 1 5 10	410
GAT GAG CTG TGG CTG GCA GTA AGA TTA GCG CAC TGC AGC AGC AAC TTT Asp Glu Leu Trp Leu Ala Val Arg Leu Ala His Cys Ser Ser Asn Phe 15 20 25	458
AGC AGC AGT AGC AGC ACA AGA ACC ACC AGC AGC AAC CAG AGG CAC AAC Ser Ser Ser Ser Ser Thr Arg Thr Thr Ser Ser Asn Gln Arg His Asn 30 35 40 45	506
CAG CAA CTC ACA ACA CTG CAA CCA AGG AGC TTA AGT ACA AAA CAC CAC Gln Gln Leu Thr Thr Leu Gln Pro Arg Ser Leu Ser Thr Lys His His 50 55 60	554
AGC AAC ATT GCA AGC GAG CAG CAC AAT AGC CAG CAA CAG GAG CCA GCA Ser Asn Ile Ala Ser Glu Gln His Asn Ser Gln Gln Gln Glu Pro Ala 65 70 75	602
TCG AAG GAC GAG GAT GTA GCC AAC CAC GGT AGA AGC AAT GAC CAG CAG Ser Lys Asp Glu Asp Val Ala Asn His Gly Arg Ser Asn Asp Gln Gln 80 85 90	650
ACG CAT CTG CAA CAG CTA GAC AGC AGC AAC ATG TTG TCG CCA AAG ACA Thr His Leu Gln Gln Leu Asp Ser Ser Asn Met Leu Ser Pro Lys Thr 95 100 105	698
GCC GCA GCA GCA ACT GCT GCC GGC GAT GAA GCA ACA ACC CAA CAA CCA Ala Ala Ala Ala Thr Ala Ala Gly Asp Glu Ala Thr Thr Gln Gln Pro 110 115 120 125	746
ACA AAC ATA AGA CTG TGT GCA CGC AAG CGA CAA CGA TTG CGT CGC CGA Thr Asn Ile Arg Leu Cys Ala Arg Lys Arg Gln Arg Leu Arg Arg Arg 130 135 140	794
CGA AAA AGA AAA CCA GCA ACC CCA AAC GAA ACA GAT ATC AAG AAA CAA Arg Lys Arg Lys Pro Ala Thr Pro Asn Glu Thr Asp Ile Lys Lys Gln 145 150 155	842
CAG CAA CTT AGC ATG CCT CCC TTC AAA ACG CGC AAA TCC ACG GAC ACC Gln Gln Leu Ser Met Pro Pro Phe Lys Thr Arg Lys Ser Thr Asp Thr 160 165 170	890
TAC AGC ACA CCA GCA GCA ACA ACC AGC TGT CCG ACA GCC ACC TAC ATG Tyr Ser Thr Pro Ala Ala Thr Thr Ser Cys Pro Thr Ala Thr Tyr Met 175 180 185	938
CAA TGT CGA GCC AGC GAC AAT GAG TTC AGT ATT CCG ATA TCG AGA CAT Gln Cys Arg Ala Ser Asp Asn Glu Phe Ser Ile Pro Ile Ser Arg His 190 195 200 205	986
GAT AGA GTA TCC ACG GCC ACA TTC GCC TGG GTG TTG CAT GTG CTG CAG Asp Arg Val Ser Thr Ala Thr Phe Ala Trp Val Leu His Val Leu Gln 210 215 220	1034
GTG CTG CTC GTG TCG CTG CAA CAG TGG CAA CTT CAC GTG CAA CAG CGA Val Leu Leu Val Ser Leu Gln Gln Trp Gln Leu His Val Gln Gln Arg 225 230 235	1082
TCG GTG CTA CTG TTC AGA AGG ATC GCA GCG AGC ACC ATC GCC TTC ATT Ser Val Leu Leu Ph Arg Arg Ile Ala Ala Ser Thr Ile Ala Phe Ile 240 245 250	1130

TCC Ser	TAT Tyr 255	TTA Leu	GGC Gly	AGC Ser	TTT Phe	GCA Ala 260	GCG Ala	CAA Gln	CTG Leu	AAA Lys	AAT Asn 265	AGC Ser	AGC Ser	AGC Ser	AGC Ser	1178
AGT Ser 270	AGC Ser	AGC Ser	AGC Ser	AAC Asn	AGC Ser 275	AGC Ser	AAC Asn	AAC Asn	AGC Ser	AGC Ser 280	ACG Thr	CAA Gln	ATA Ile	TTA Leu	AAC Asn 285	1226
GGA Gly	CTT Leu	AAT Asn	AAA Lys	CAC His 290	TCA Ser	TGG Trp	ATA Ile	TTT Phe	TTA Leu 295	TTG Leu	ATA Ile	TAT Tyr	TTG Leu	AAT Asn 300	TTA Leu	1274
TCT Ser	GCT Ala	AAA Lys	GTT Val 305	TGC Cys	CTA Leu	GCA Ala	GGA Gly	TAT Tyr 310	CAT His	GAA Glu	AAG Lys	AGA Arg	CTG Leu 315	TTA Leu	CAC His	1322
GAT Asp	CTT Leu	TTG Leu 320	GAT Asp	CCT Pro	TAT Tyr	AAT Asn	ACA Thr 325	CTA Leu	GAA Glu	CGT Arg	CCC Pro	GTT Val 330	CTC Leu	AAT Asn	GAA Glu	1370
TCG Ser 335	GAC Asp	CCG Pro	TTA Leu	CAA Gln	TTA Leu	AGC Ser 340	TTT Phe	GGT Gly	TTA Leu	ACT Thr	TTA Leu 345	ATG Met	CAA Gln	ATT Ile	ATC Ile	1418
GAT Asp 350	GTG Val	GAC Asp	GAG Glu	AAA Lys	AAT Asn 355	CAA Gln	TTG Leu	CTA Leu	GTC Val	ACT Thr 360	AAT Asn	GTG Val	TGG Trp	TTA Leu	AAA Lys 365	1466
CTG Leu	GAG Glu	TGG Trp	AAC Asn	GAC Asp 370	ATG Met	AAT Asn	CTC Leu	CGC Arg	TGG Trp 375	AAC Asn	ACC Thr	TCC Ser	GAC Asp	TAT Tyr 380	GGC Gly	1514
GGA Gly	GTT Val	AAG Lys	GAT Asp 385	CTG Leu	CGA Arg	ATA Ile	CCG Pro	CCG Pro 390	CAT His	CGC Arg	ATC Ile	TGG Trp	AAG Lys 395	CCG Pro	GAC Asp	1562
GTG Val	CTG Leu	ATG Met 400	TAC Tyr	AAC Asn	AGT Ser	GCG Ala	GAT Asp 405	GAG Glu	GGA Gly	TTT Phe	GAC Asp	GGC Gly 410	ACC Thr	TAC Tyr	CAG Gln	1610
ACG Thr 415	AAC Asn	GTG Val	GTG Val	GTG Val	CGG Arg	AAC Asn 420	AAC Asn	GGC Gly	TCG Ser	TGT Cys	CTA Leu 425	TAC Tyr	GTT Val	CCG Pro	CCG Pro	1658
GGG Gly 430	ATC Ile	TTC Phe	AAG Lys	TCG Ser	ACG Thr 435	TGC Cys	AAG Lys	ATC Ile	GAC Asp 440	ATC Ile	ACG Thr	TGG Trp	TTC Phe	CCC Pro	TTC Phe 445	1706
GAT Asp	GAC Asp	CAG Gln	CGG Arg	TGC Cys 450	GAG Glu	ATG Met	AAG Lys	TTC Phe	GGC Gly 455	AGT Ser	TGG Trp	ACC Thr	TAC Tyr	GAC Asp 460	GGA Gly	1754
TTC Phe	CAG Gln	CTG Leu	GAT Asp 465	TTA Leu	CAA Gln	TTA Leu	CAA Gln	GAT Asp 470	GAA Glu	ACT Thr	GGC Gly	GGT Gly	GAT Asp 475	ATC Ile	AGC Ser	1802
AGT Ser	TAC Tyr	GTG Val 480	CTC Leu	AAC Asn	GGC Gly	GAG Glu	TGG Trp 485	GAA Glu	CTA Leu	CTG Leu	GGT Gly	GTG Val 490	CCC Pro	GGC Gly	AAA Lys	1850
CGT Arg	AAC Asn 495	GAG Glu	ATC Ile	TAT Tyr	TAC Tyr	AAC Asn 500	TGC Cys	TGC Cys	CCG Pro	GAA Glu	CCC Pro 505	TAT Tyr	ATA Ile	GAC Asp	ATC Ile	1898
ACC Thr 510	TTC Phe	GCC Ala	ATC Ile	ATC Ile	ATC Ile 515	CGC Arg	CGA Arg	CGA Arg	ACA Thr	CTG Leu 520	TAC Tyr	TAT Tyr	TTC Phe	TTC Ph	AAC Asn 525	1946

CTG ATC ATA CCT TGT GTA CTG ATT GCC TCC ATG GCC TTG CTC GGA TTC Leu Ile Ile Pro Cys Val Leu Ile Ala Ser Met Ala Leu Leu Gly Phe 530 535 540	1994
ACC CTG CCG CCA GAT TCG GGT GAA AAA TTA TCG CTG GGT GTT ACC ATC Thr Leu Pro Pro Asp Ser Gly Glu Lys Leu Ser Leu Gly Val Thr Ile 545 550 555	2042
TTG CTC TCG CTG ACC GTG TTT CTG AAT ATG GTT GCC GAG ACA ATG CCG Leu Leu Ser Leu Thr Val Phe Leu Asn Met Val Ala Glu Thr Met Pro 560 565 570	2090
GCT ACT TCC GAT GCG GTG CCA TTG TGG ATA CGC ATC GTG TTT TTG TGC Ala Thr Ser Asp Ala Val Pro Leu Trp Ile Arg Ile Val Phe Leu Cys 575 580 585	2138
TGG CTG CCA TGG ATA TTG CGA ATG AGT CGC CCA GGA CGA CCG CTG ATC Trp Leu Pro Trp Ile Leu Arg Met Ser Arg Pro Gly Arg Pro Leu Ile 590 595 600 605	2186
CTA GAG TTC CCG ACC ACG CCC TGT TCG GAC ACA TCC TCC GAG CGG AAG Leu Glu Phe Pro Thr Thr Pro Cys Ser Asp Thr Ser Ser Glu Arg Lys 610 615 620	2234
CAC CAG ATA CTC TCC GAC GTT GAG CTG AAA GAG CGC TCG TCG AAA TCG His Gln Ile Leu Ser Asp Val Glu Leu Lys Glu Arg Ser Ser Lys Ser 625 630 635	2282
CTG CTG GCC AAC GTA CTA GAC ATC GAT GAT GAC TTC CGG CAC AAT TGT Leu Leu Ala Asn Val Leu Asp Ile Asp Asp Asp Phe Arg His Asn Cys 640 645 650	2330
CGC CCC ATG ACG CCC GGC GGA ACA CTG CCA CAC AAC CCG GCT TTC TAT Arg Pro Met Thr Pro Gly Gly Thr Leu Pro His Asn Pro Ala Phe Tyr 655 660 665	2378
CGC ACG GTT TAT GGA CAA GGC GAC GAT GGC AGC ATT GGG CCA ATT GGC Arg Thr Val Tyr Gly Gln Gly Asp Asp Gly Ser Ile Gly Pro Ile Gly 670 675 680 685	2426
AGC ACC CGA ATG CCG GAT GCG GTC ACC CAT CAT ACG TGC ATC AAA TCA Ser Thr Arg Met Pro Asp Ala Val Thr His His Thr Cys Ile Lys Ser 690 695 700	2474
TCA ACT GAA TAT GAA TTA GGT TTA ATC TTA AAG GAA ATT CGC TTT ATA Ser Thr Glu Tyr Glu Leu Gly Leu Ile Leu Lys Glu Ile Arg Phe Ile 705 710 715	2522
ACT GAT CAG CTA CGT AAA GAT GAC GAG TGC AAT GAC ATT GCC AAT GAT Thr Asp Gln Leu Arg Lys Asp Asp Glu Cys Asn Asp Ile Ala Asn Asp 720 725 730	2570
TGG AAA TTT GCA GCT ATG GTC GTT GAC AGA CTG TGC CTT ATC ATA TTC Trp Lys Phe Ala Ala Met Val Val Asp Arg Leu Cys Leu Ile Ile Phe 735 740 745	2618
ACA ATG TTC GCA ATA TTA GCC ACA ATA GCT GTA CTA CTA TCG GCA CCA Thr Met Phe Ala Ile Leu Ala Thr Ile Ala Val Leu Leu Ser Ala Pro 750 755 760 765	2666
CAT ATT ATT GTC TCG TAGCCATATG GGCGAGGTGG TTATTGTTAT TGTTTTTATT His Ile Ile Val Ser 770	2721
ATAAAATCAA TTTGTTAATT ATTAAATTAA TAACGAACT CTTTAAGTAA ATTAAACTA	2781
AAAAAGACACT AAAAAAGCAC AAAAAAATAG GAAATACAT GATAAAACCC ATGAACTAAA	2841

TAATACATCC AAGAAAAACC AAAACAAAAA AAAAAAAAAA AAAAA

2886

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 770 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) TYPE OF MOLECULE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

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Met Lys Asn Ala Gln Leu Lys Leu Thr Glu Val Asp Asp Asp Glu Leu
 1          5          10          15
Trp Leu Ala Val Arg Leu Ala His Cys Ser Ser Asn Phe Ser Ser Ser
          20          25          30
Ser Ser Thr Arg Thr Thr Ser Ser Asn Gln Arg His Asn Gln Gln Leu
          35          40          45
Thr Thr Leu Gln Pro Arg Ser Leu Ser Thr Lys His His Ser Asn Ile
          50          55          60
Ala Ser Glu Gln His Asn Ser Gln Gln Gln Glu Pro Ala Ser Lys Asp
          65          70          75          80
Glu Asp Val Ala Asn His Gly Arg Ser Asn Asp Gln Gln Thr His Leu
          85          90          95
Gln Gln Leu Asp Ser Ser Asn Met Leu Ser Pro Lys Thr Ala Ala Ala
          100          105          110
Ala Thr Ala Ala Gly Asp Glu Ala Thr Thr Gln Gln Pro Thr Asn Ile
          115          120          125
Arg Leu Cys Ala Arg Lys Arg Gln Arg Leu Arg Arg Arg Arg Lys Arg
          130          135          140
Lys Pro Ala Thr Pro Asn Glu Thr Asp Ile Lys Lys Gln Gln Gln Leu
          145          150          155          160
Ser Met Pro Pro Phe Lys Thr Arg Lys Ser Thr Asp Thr Tyr Ser Thr
          165          170          175
Pro Ala Ala Thr Thr Ser Cys Pro Thr Ala Thr Tyr Met Gln Cys Arg
          180          185          190
Ala Ser Asp Asn Glu Phe Ser Ile Pro Ile Ser Arg His Asp Arg Val
          195          200          205
Ser Thr Ala Thr Phe Ala Trp Val Leu His Val Leu Gln Val Leu Leu
          210          215          220
Val Ser Leu Gln Gln Trp Gln Leu His Val Gln Gln Arg Ser Val Leu
          225          230          235          240
Leu Phe Arg Arg Ile Ala Ala Ser Thr Ile Ala Phe Ile Ser Tyr Leu
          245          250          255
Gly Ser Phe Ala Ala Gln Leu Lys Asn Ser Ser Ser Ser Ser Ser Ser
          260          265          270
Ser Asn S r Ser Asn Asn Ser Ser Thr Gln Ile Leu Asn Gly Leu Asn
          275          280          285

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Lys	His	Ser	Trp	Ile	Phe	Leu	Leu	Ile	Tyr	Leu	Asn	Leu	Ser	Ala	Lys
	290					295					300				
Val	Cys	Leu	Ala	Gly	Tyr	His	Glu	Lys	Arg	Leu	Leu	His	Asp	Leu	Leu
305					310					315					320
Asp	Pro	Tyr	Asn	Thr	Leu	Glu	Arg	Pro	Val	Leu	Asn	Glu	Ser	Asp	Pro
				325					330					335	
Leu	Gln	Leu	Ser	Phe	Gly	Leu	Thr	Leu	Met	Gln	Ile	Ile	Asp	Val	Asp
			340					345					350		
Glu	Lys	Asn	Gln	Leu	Leu	Val	Thr	Asn	Val	Trp	Leu	Lys	Leu	Glu	Trp
		355					360					365			
Asn	Asp	Met	Asn	Leu	Arg	Trp	Asn	Thr	Ser	Asp	Tyr	Gly	Gly	Val	Lys
	370					375					380				
Asp	Leu	Arg	Ile	Pro	Pro	His	Arg	Ile	Trp	Lys	Pro	Asp	Val	Leu	Met
385					390					395					400
Tyr	Asn	Ser	Ala	Asp	Glu	Gly	Phe	Asp	Gly	Thr	Tyr	Gln	Thr	Asn	Val
				405					410					415	
Val	Val	Arg	Asn	Asn	Gly	Ser	Cys	Leu	Tyr	Val	Pro	Pro	Gly	Ile	Phe
			420					425					430		
Lys	Ser	Thr	Cys	Lys	Ile	Asp	Ile	Thr	Trp	Phe	Pro	Phe	Asp	Asp	Gln
		435					440					445			
Arg	Cys	Glu	Met	Lys	Phe	Gly	Ser	Trp	Thr	Tyr	Asp	Gly	Phe	Gln	Leu
	450					455					460				
Asp	Leu	Gln	Leu	Gln	Asp	Glu	Thr	Gly	Gly	Asp	Ile	Ser	Ser	Tyr	Val
465					470					475					480
Leu	Asn	Gly	Glu	Trp	Glu	Leu	Leu	Gly	Val	Pro	Gly	Lys	Arg	Asn	Glu
				485					490					495	
Ile	Tyr	Tyr	Asn	Cys	Cys	Pro	Glu	Pro	Tyr	Ile	Asp	Ile	Thr	Phe	Ala
			500					505					510		
Ile	Ile	Ile	Arg	Arg	Arg	Thr	Leu	Tyr	Tyr	Phe	Phe	Asn	Leu	Ile	Ile
		515					520					525			
Pro	Cys	Val	Leu	Ile	Ala	Ser	Met	Ala	Leu	Leu	Gly	Phe	Thr	Leu	Pro
	530					535					540				
Pro	Asp	Ser	Gly	Glu	Lys	Leu	Ser	Leu	Gly	Val	Thr	Ile	Leu	Leu	Ser
545					550					555					560
Leu	Thr	Val	Phe	Leu	Asn	Met	Val	Ala	Glu	Thr	Met	Pro	Ala	Thr	Ser
				565					570					575	
Asp	Ala	Val	Pro	Leu	Trp	Ile	Arg	Ile	Val	Phe	Leu	Cys	Trp	Leu	Pro
			580					585					590		
Trp	Ile	Leu	Arg	Met	Ser	Arg	Pro	Gly	Arg	Pro	Leu	Ile	Leu	Glu	Phe
	595						600					605			
Pro	Thr	Thr	Pro	Cys	Ser	Asp	Thr	Ser	Ser	Glu	Arg	Lys	His	Gln	Ile
	610					615					620				
Leu	Ser	Asp	Val	Glu	Leu	Lys	Glu	Arg	Ser	Ser	Lys	Ser	Leu	Leu	Ala
625					630					635					640
Asn	Val	Leu	Asp	Ile	Asp	Asp	Asp	Phe	Arg	His	Asn	Cys	Arg	Pro	Met

645								650				655			
Thr	Pro	Gly	Gly 660	Thr	Leu	Pro	His	Asn 665	Pro	Ala	Phe	Tyr	Arg 670	Thr	Val
Tyr	Gly	Gln 675	Gly	Asp	Asp	Gly	Ser 680	Ile	Gly	Pro	Ile	Gly 685	Ser	Thr	Arg
Met	Pro 690	Asp	Ala	Val	Thr	His 695	His	Thr	Cys	Ile	Lys 700	Ser	Ser	Thr	Glu
Tyr 705	Glu	Leu	Gly	Leu	Ile 710	Leu	Lys	Glu	Ile	Arg 715	Phe	Ile	Thr	Asp	Gln 720
Leu	Arg	Lys	Asp	Asp 725	Glu	Cys	Asn	Asp	Ile 730	Ala	Asn	Asp	Trp	Lys 735	Phe
Ala	Ala	Met	Val 740	Val	Asp	Arg	Leu	Cys 745	Leu	Ile	Ile	Phe	Thr 750	Met	Phe
Ala	Ile	Leu 755	Ala	Thr	Ile	Ala	Val 760	Leu	Leu	Ser	Ala	Pro 765	His	Ile	Ile
Val	Ser 770														

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 3700 base pairs
(B) TYPE: nucleotide
(C) STRANDEDNESS: double strand
(D) TOPOLOGY: linear

(ii) TYPE OF MOLECULE: cDNA of mRNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: *Heliothis virescens*

(vii) IMMEDIATE SOURCE:

- (B) CLONE(S) : Hva7-1

(ix) FEATURES:

- (A) NAME/KEY: CDS
(B) POSITION: 335..1822

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

GGCACGAGCC	GCTGCCCCAC	GGTCGGCCGC	ACTCCGCTGA	ACAACAATGC	TCAAAAACAC	60
GCCGTGACTC	CACACACATC	CCCTCGGCGC	AGTAGGCGAT	GTTTGAGGAT	CGGACGGCAC	120
GCGTGGCCGT	CGGCGAGCGG	TCGTGAACAA	GTTGCATACA	TATGAAAACC	GTA AAAAGAT	180
TGAATTTTAA	GCCGATCGTG	TTCGATAGAT	CCTAATAGAG	AAGCGGGAGT	GCGGCGTTTG	240
GTAGGCGGGG	GTCGAGTCGC	GCGGTCGGGG	GAAATGGCGC	GGCGCGGGGC	GCGGCGGGCG	300
GCGGCGCGCG	GCGGCGGGCG	GTCGCGGGCG	TGAC	ATG GGC GGG CGG GCG CGC		352
			Met Gly Gly Arg	Ala Arg		
				775		

CGC Arg	TCG Ser	CAC His	TTG Leu 780	GCG Ala	GCG Ala	CCC Pro	GCG Ala	GGC Gly 785	CTG Leu	CTG Leu	CTG Leu	CTG Leu	CTG Leu	TGC Cys 790	CTG Leu	400
CTC Leu	TGG Trp	CCG Pro 795	AGG Arg	GGG Gly	GCA Ala	CGC Arg	TGC Cys 800	GGG Gly	TAC Tyr	CAC His	GAG Glu	AAG Lys 805	CGG Arg	CTA Leu	CTG Leu	448
CAC His	CAC His 810	CTA Leu	TTG Leu	GAC Asp	CAC His	TAC Tyr 815	AAC Asn	GTA Val	CTG Leu	GAG Glu	AGG Arg 820	CCC Pro	GTC Val	GTC Val	AAC Asn	496
GAG Glu 825	AGC Ser	GAC Asp	CCG Pro	CTG Leu	CAG Gln 830	CTC Leu	TCC Ser	TTC Phe	GGC Gly	CTC Leu 835	ACG Thr	CTC Leu	ATG Met	CAG Gln	ATC Ile 840	544
ATC Ile	GAC Asp	GTG Val	GAC Asp	GAG Glu 845	AAG Lys	AAC Asn	CAG Gln	CTT Leu	TTA Leu 850	ATA Ile	ACA Thr	AAC Asn	ATC Ile	TGG Trp 855	CTA Leu	592
AAA Lys	CTA Leu	GAG Glu	TGG Trp 860	AAT Asn	GAT Asp	ATG Met	AAC Asn	TTG Leu 865	AGG Arg	TGG Trp	AAC Asn	ACT Thr	TCA Ser 870	GAT Asp	TTC Phe	640
GGC Gly	GGG Gly	GTC Val 875	AAA Lys	GAT Asp	TTA Leu	AGA Arg	GTG Val 880	CCA Pro	CCC Pro	CAC His	AGA Arg	CTA Leu 885	TGG Trp	AAA Lys	CCA Pro	688
GAC Asp	GTC Val 890	CTT Leu	ATG Met	TAC Tyr	AAC Asn	AGC Ser 895	GCG Ala	GAC Asp	GAA Glu	GGG Gly	TTC Phe 900	GAC Asp	AGC Ser	ACG Thr	TAT Tyr	736
CCA Pro 905	ACG Thr	AAC Asn	GTG Val	GTG Val 910	GTG Val	CGG Arg	AAC Asn	AAC Asn	GGC Gly	TCG Ser 915	TGT Cys	CTG Leu	TAC Tyr	GTG Val	CCG Pro 920	784
CCC Pro	GGC Gly	ATC Ile	TTC Phe	AAG Lys 925	AGC Ser	ACC Thr	TGC Cys	AAG Lys	ATC Ile 930	GAC Asp	ATC Ile	ACC Thr	TGG Trp	TTC Phe	CCC Pro 935	832
TTC Phe	GAC Asp	GAC Asp	CAA Gln 940	CGA Arg	TGC Cys	GAG Glu	ATG Met	AAG Lys 945	TTT Phe	GGC Gly	AGC Ser	TGG Trp	ACT Thr 950	TAT Tyr	GAT Asp	880
GGT Gly	TAT Tyr	CAG Gln 955	TTG Leu	GAT Asp	CTA Leu	CAA Gln	CTA Leu 960	CAG Gln	GAT Asp	GAA Glu	GGG Gly	GGC Gly 965	GGA Gly	GAT Asp	ATA Ile	928
AGC Ser	AGT Ser	TTT Phe 970	GTC Val	ACG Thr	AAT Asn	GGC Gly 975	GAA Glu	TGG Trp	GAG Glu	TTA Leu	ATA Ile 980	GGA Gly	GTC Val	CCC Pro	GGC Gly	976
AAG Lys 985	CGC Arg	AAC Asn	GAG Glu	ATC Ile	TAC Tyr 990	TAC Tyr	AAC Asn	TGT Cys	TGT Cys	CCG Pro 995	GAG Glu	CCA Pro	TAC Tyr	ATC Ile	GAC Asp 1000	1024
ATC Ile	ACG Thr	TTT Phe	GCG Ala 1005	GTG Val	GTG Val	ATC Ile	CGG Arg	AGG Arg	AAA Lys 1010	ACG Thr	CTC Leu	TAC Tyr	TAC Tyr	TTC Phe	TTC Phe 1015	1072
AAT Asn	CTG Leu	ATC Ile	GTG Val 1020	CCC Pro	TGC Cys	GTG Val	CTC Leu	ATC Ile 1025	GCC Ala	TCC Ser	ATG Met	GCT Ala	CTA Leu 1030	TTG Leu	GGG Gly	1120
TTC Phe	ACC Thr	TTG Leu 1035	CCT Pro	CCA Pro	GAC Asp	TCC Ser	GGA Gly 1040	GAA Glu	AAG Lys	TTG Leu	TCT Ser	TTA Leu 1045	GGT Gly	GTG Val	ACG Thr	1168

ATA TTA CTG TCG TTG ACG GTG TTC CTC AAC ATG GTG GCG GAG ACG ATG Ile Leu Leu Ser Leu Thr Val Phe Leu Asn Met Val Ala Glu Thr Met 1050 1055 1060	1216
CCA GCG ACG TCG GAC GCC GTG CCC TTG CTC GGC ACC TAC TTC AAC TGC Pro Ala Thr Ser Asp Ala Val Pro Leu Leu Gly Thr Tyr Phe Asn Cys 1065 1070 1075 1080	1264
ATC ATG TTC ATG GTG GCT TCC TCC GTC GTC TCC ACC ATA CTG ATC CTC Ile Met Phe Met Val Ala Ser Ser Val Val Ser Thr Ile Leu Ile Leu 1085 1090 1095	1312
AAC TAC CAC CAC CGG CAC GCA GAC ACT CAC GAA ATG AGT GAT TGG ATT Asn Tyr His His Arg His Ala Asp Thr His Glu Met Ser Asp Trp Ile 1100 1105 1110	1360
CGT TGC GTG TTC CTT TAT TGG CTG CCG TGG GTG CTG CGC ATG TCA CGG Arg Cys Val Phe Leu Tyr Trp Leu Pro Trp Val Leu Arg Met Ser Arg 1115 1120 1125	1408
CCC GGC TCG GCG ACG ACG CCG CCG CCG GCG CGC GTA CCT CCG CCG CCG Pro Gly Ser Ala Thr Thr Pro Pro Pro Ala Arg Val Pro Pro Pro Pro 1130 1135 1140	1456
GAC CTG GAG CTG CGC GAG CGC TCC TCC AAG TCG CTC CTA GCG AAC GTG Asp Leu Glu Leu Arg Glu Arg Ser Ser Lys Ser Leu Leu Ala Asn Val 1145 1150 1155 1160	1504
CTC GAC ATC GAT GAC GAC TTC CGC CAC CCG CAA GCG CAG CAG CCG CAA Leu Asp Ile Asp Asp Asp Phe Arg His Pro Gln Ala Gln Gln Pro Gln 1165 1170 1175	1552
TGC TGC CGA TAC TAC AGG GGG GGT GAG GAG AAT GGC GCG GGG TTG GCG Cys Cys Arg Tyr Tyr Arg Gly Gly Glu Glu Asn Gly Ala Gly Leu Ala 1180 1185 1190	1600
GCG CAC AGT TGC TTC GGT GTC GAC TAC GAG CTC TCC CTC ATT CTG AAG Ala His Ser Cys Phe Gly Val Asp Tyr Glu Leu Ser Leu Ile Leu Lys 1195 1200 1205	1648
GAG ATT AGA GTC ATC ACA GAT CAG ATG CGC AAG GAC GAC GAA GAT GCG Glu Ile Arg Val Ile Thr Asp Gln Met Arg Lys Asp Asp Glu Asp Ala 1210 1215 1220	1696
GAC ATT TCG CGC GAC TGG AAG TTC GCC GCC ATG GTC GTG GAC AGA CTG Asp Ile Ser Arg Asp Trp Lys Phe Ala Ala Met Val Val Asp Arg Leu 1225 1230 1235 1240	1744
TGC CTT ATT ATC TTT ACC CTG TTC ACA ATC ATC GCC ACG CTA GCC GTG Cys Leu Ile Ile Phe Thr Leu Phe Thr Ile Ile Ala Thr Leu Ala Val 1245 1250 1255	1792
CTG CTG TCC GCG CCA CAC ATC ATG GTG TCG TAGCGACCCG CCCGCTTGCG Leu Leu Ser Ala Pro His Ile Met Val Ser 1260 1265	1842
GATACGCATG CGAAAAGTTC TGTGATACCG CGAATATTTG TTAAGTTGTG ATGAGCGAAG	1902
TGGCGCGGAC GGTGACGCCG CGGCGTCGGA GTTGCCGCCG CCTGCCTCGC CGCCCGCGCC	1962
CCCCTGTAGA CATAAGTTAC CGCTGACTGC CAACCCTGTA CGTTCAACAA ATAAGTCCCC	2022
ATCCGACTAA CGTCTTTTAT CCCCTTGAAA AATTCAGCGA TTGTGTACCC CTTTCTTCCA	2082
AGAATACAAT GACAAATGGT CGTCACGCTC AGTGGAATCA ATCCCGTACT CTTGCCCCGA	2142
TATTTCCCTT AGGGTATGTC ACGAGTTTGA ATGAGCGGTT CCGTATCAGA CGTTCCGTCC	2202

CCGGAACGGT	CGTCCCCTGC	GATAAAGTGG	CAGTACGTGC	TATACAGGCA	CTTAAGGCCG	2262
CCACGCCACG	GCGCCGCGGT	GCGCTCGGGC	CGCGAACCCG	CGACCCTCAC	CGCTGCAAGT	2322
GGCCACCCAC	TAGACAAGAC	TGCGGCAGAA	AATATTTGCA	CAAAAACGTC	TTCCTTCTTA	2382
CCGATGAACG	ACCTGATTCT	CATTTAAAAA	TAAACTTTGT	TAGAACTTCT	TCGATTCTTG	2442
AAATCTATTG	TACAGTTTAG	AGTTTGGGCG	GTGAAACAAT	GGCCCTTTGT	TTCCTTCTTG	2502
TTTCGATTCCA	TGAATCGTGG	TTATAATCCC	TAGTTTTATT	TTCGGATATA	TTTGTGTCAG	2562
TAGCTAGTAT	AGAACTTTAC	AAACAATGTT	GATTCAATTG	GTACAGGTTG	TGATATGCCT	2622
CGTTGTGAAC	GGGTCCGATA	TTGTTATAAA	TGGTAAAATA	CCCATGGCTA	TAGCTTAATA	2682
AATCGTTCGT	TAAAAGTTGT	AGTTAAACAA	ATATTATTTT	AATAAAGTCA	TATCTGGGTC	2742
TTCCGGAACG	ACTTTTACAA	ATAATTAAAT	TACATATTAA	TATCACGTTT	GTACTTCTTT	2802
CCATACAGTT	ACAGTAATTC	GTATGCTGAA	AATAATATTA	GCTTGTAATA	TTTTCTTCTT	2862
CGAAAATTTA	TTCAAACAGA	TGCGACCATC	GTTTCAAACA	TTTACATGTA	ATATAGAACT	2922
CATTTTATAA	GATATACAAC	ATTTTATAAG	TACAAGAAGT	TGTAACATGA	ACCGGTTTTT	2982
CGTTACATAG	AGGGTATAAC	ACAAAGGTGC	CTACATATTG	ACAGATGCGA	AGCACGATCA	3042
GTTGATAAGC	ACAGGTACAC	TATATCCTGA	CATCCGACAG	TCCTGCCGCT	CGTCTGCCAC	3102
ACTCGGAAAC	ATTCGACAGT	TCAGTTTACT	GCTCCGCCAT	CATCGATTGT	TAAGTTTGTT	3162
GTTCTAACTC	ATCGCATTCA	TTTCATTCAA	AAACATTGTA	AACCTCTCAA	GGGGAAAACG	3222
TGTTGTAAAC	AGTGAGAGTG	CGCGGGTACA	ACCGACACGC	GAATGTACCC	TCGCAAGGCT	3282
CCTGTAATGT	TTTCCTCTTC	CGAGGTGTTG	CTGAGAGTAA	TCTTAGACGG	TCCGATGGAA	3342
GTTGCGGACC	GGATATGATT	ACAAGTCAAT	GTTTTTAAGT	CATCCGTTTA	TTTATTGTTA	3402
TATCTTCTTA	CCATTCGCTA	GAGGTTGTGT	GACGACCCGG	ACGGTGGGCG	CCGCAACCCG	3462
CACACGCGGG	GTTCCATCTT	TGTATTAGAT	GGAAGTTGTG	CGGCATCTCT	CCGTCGGCAA	3522
TGGGACAACC	CGTTGTCCCC	AACATTTGTT	CAATTGTTAG	GGTTAACTCT	GAATTGCACT	3582
TTGTTTATTA	AATATAAACG	AATGAAACAA	AAAAAAAAAA	AAAAAACTCG	AGAGTACTTC	3642
TAGAGCGGCC	GCGGGCCCAT	CGATTTTCCA	CCCGGTGGG	GTACCAGTAA	GTGTACCC	3700

(2) INFORMATION FOR SEQ ID NO: 4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 496 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) TYPE OF MOLECULE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4

Met	Gly	Gly	Arg	Ala	Arg	Arg	Ser	His	Leu	Ala	Ala	Pro	Ala	Gly	Leu
1				5					10					15	
Leu	Leu	Leu	Leu	Cys	Leu	Leu	Trp	Pro	Arg	Gly	Ala	Arg	Cys	Gly	Tyr
			20					25					30		

His Glu Lys Arg Leu Leu His His Leu Leu Asp His Tyr Asn Val Leu
 35 40 45
 Glu Arg Pro Val Val Asn Glu Ser Asp Pro Leu Gln Leu Ser Phe Gly
 50 55 60
 Leu Thr Leu Met Gln Ile Ile Asp Val Asp Glu Lys Asn Gln Leu Leu
 65 70 75 80
 Ile Thr Asn Ile Trp Leu Lys Leu Glu Trp Asn Asp Met Asn Leu Arg
 85 90 95
 Trp Asn Thr Ser Asp Phe Gly Gly Val Lys Asp Leu Arg Val Pro Pro
 100 105 110
 His Arg Leu Trp Lys Pro Asp Val Leu Met Tyr Asn Ser Ala Asp Glu
 115 120 125
 Gly Phe Asp Ser Thr Tyr Pro Thr Asn Val Val Val Arg Asn Asn Gly
 130 135 140
 Ser Cys Leu Tyr Val Pro Gly Ile Phe Lys Ser Thr Cys Lys Ile
 145 150 155 160
 Asp Ile Thr Trp Phe Pro Phe Asp Asp Gln Arg Cys Glu Met Lys Phe
 165 170 175
 Gly Ser Trp Thr Tyr Asp Gly Tyr Gln Leu Asp Leu Gln Leu Gln Asp
 180 185 190
 Glu Gly Gly Gly Asp Ile Ser Ser Phe Val Thr Asn Gly Glu Trp Glu
 195 200 205
 Leu Ile Gly Val Pro Gly Lys Arg Asn Glu Ile Tyr Tyr Asn Cys Cys
 210 215 220
 Pro Glu Pro Tyr Ile Asp Ile Thr Phe Ala Val Val Ile Arg Arg Lys
 225 230 235 240
 Thr Leu Tyr Tyr Phe Phe Asn Leu Ile Val Pro Cys Val Leu Ile Ala
 245 250 255
 Ser Met Ala Leu Leu Gly Phe Thr Leu Pro Pro Asp Ser Gly Glu Lys
 260 265 270
 Leu Ser Leu Gly Val Thr Ile Leu Leu Ser Leu Thr Val Phe Leu Asn
 275 280 285
 Met Val Ala Glu Thr Met Pro Ala Thr Ser Asp Ala Val Pro Leu Leu
 290 295 300
 Gly Thr Tyr Phe Asn Cys Ile Met Phe Met Val Ala Ser Ser Val Val
 305 310 315 320
 Ser Thr Ile Leu Ile Leu Asn Tyr His His Arg His Ala Asp Thr His
 325 330 335
 Glu Met Ser Asp Trp Ile Arg Cys Val Phe Leu Tyr Trp Leu Pro Trp
 340 345 350
 Val Leu Arg Met Ser Arg Pro Gly Ser Ala Thr Thr Pro Pro Pro Ala
 355 360 365
 Arg Val Pro Pro Pro Pro Asp Leu Glu Leu Arg Glu Arg Ser Ser Lys
 370 375 380
 Ser Leu Leu Ala Asn Val Leu Asp Ile Asp Asp Asp Phe Arg His Pro

385		390		395		400									
Gln	Ala	Gln	Gln	Pro	Gln	Cys	Cys	Arg	Tyr	Tyr	Arg	Gly	Gly	Glu	Glu
				405					410					415	
Asn	Gly	Ala	Gly	Leu	Ala	Ala	His	Ser	Cys	Phe	Gly	Val	Asp	Tyr	Glu
			420					425					430		
Leu	Ser	Leu	Ile	Leu	Lys	Glu	Ile	Arg	Val	Ile	Thr	Asp	Gln	Met	Arg
		435					440					445			
Lys	Asp	Asp	Glu	Asp	Ala	Asp	Ile	Ser	Arg	Asp	Trp	Lys	Phe	Ala	Ala
	450					455					460				
Met	Val	Val	Asp	Arg	Leu	Cys	Leu	Ile	Ile	Phe	Thr	Leu	Phe	Thr	Ile
465					470					475					480
Ile	Ala	Thr	Leu	Ala	Val	Leu	Leu	Ser	Ala	Pro	His	Ile	Met	Val	Ser
			485					490						495	

(2) INFORMATION FOR SEQ ID NO: 5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 3109 base pairs
- (B) TYPE: nucleotide
- (C) STRANDEDNESS: double strand
- (D) TOPOLOGY: linear

(ii) TYPE OF MOLECULE: cDNA of mRNA

(iii) HYPOTHETICAL: NO

(iv) ANTISENSE: NO

(vi) ORIGINAL SOURCE:

(A) ORGANISM: *Heliothis virescens*

(vii) IMMEDIATE SOURCE:

(B) CLONE(S): Hva7-2

(ix) FEATURES:

- (A) NAME/KEY: CDS
- (B) POSITION: 95..1597

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

GGCACGAGCC	GGCCGCACGT	TGTCCCAGGC	CGCATGAGCG	CGCCGGCGTG	CTAGCGCAGC	60
GTGCGCGGGT	GTGGTATGCC	CGCGCGTCGC	CGCT ATG GCC	CCT ATG TTG	GCG	112
			Met Ala	Pro Met	Leu Ala	
				500		
GCC TTG GCG	CTG CTG GCT	TTG CTG CCC	GTA TCG GAG	CAA GGT	CCT CAC	160
Ala Leu Ala	Leu Leu Ala	Leu Leu Pro	Val Ser Glu	Gln Gly	Pro His	
	505		510		515	
GAG AAG AGA	CTC CTG AAC	GCG TTG CTG	GCG AAC TAC	AAC ACC	CTG GAG	208
Glu Lys Arg	Leu Leu Asn	Ala Leu Leu	Ala Asn Tyr	Asn Thr	Leu Glu	
	520		525		530	
CGA CCG GTG	GCC AAC GAG	AGC GAA CCG	CTA GAG	GTC AGG	TTC GGC	256
Arg Pro Val	Ala Asn Glu	Ser Glu Pro	Leu Glu	Val Arg	Phe Gly	
	535		540		550	
ACC TTG CAG	CAA ATC ATT	GAC GTG	GAC GAG	AAG AAT	CAA CTA	304
					CTT ATA	

Thr	Leu	Gln	Gln	Ile 555	Ile	Asp	Val	Asp	Glu 560	Lys	Asn	Gln	Leu	Leu 565	Ile		
ACC	AAT	ATA	TGG	CTG	TCG	TTG	GAG	TGG	AAT	GAC	TAC	AAC	CTG	AGG	TGG	352	
Thr	Asn	Ile	Trp 570	Leu	Ser	Leu	Glu	Trp 575	Asn	Asp	Tyr	Asn	Leu 580	Arg	Trp		
AAC	GAC	AGC	GAG	TAT	GGC	GGG	GTC	AAG	GAC	CTC	AGG	ATC	ACG	CCC	AAC	400	
Asn	Asp	Ser 585	Glu	Tyr	Gly	Gly	Val 590	Lys	Asp	Leu	Arg	Ile 595	Thr	Pro	Asn		
AAG	TTG	TGG	AAG	CCG	GAC	GTC	CTT	ATG	TAT	AAT	AGT	GCT	GAC	GAG	GGT	448	
Lys	Leu 600	Trp	Lys	Pro	Asp	Val 605	Leu	Met	Tyr	Asn	Ser 610	Ala	Asp	Glu	Gly		
TTT	GAC	GGG	ACC	TAC	CAG	ACC	AAC	GTG	GTG	GTC	AGA	AGC	GGC	GGC	AGT	496	
Phe	Asp	Gly	Thr	Tyr	Gln 620	Thr	Asn	Val	Val	Val 625	Arg	Ser	Gly	Gly	Ser 630		
TGC	CTG	TAC	GTG	CCA	CCT	GGC	ATA	TTC	AAG	AGC	ACA	TGC	AAG	ATG	GAC	544	
Cys	Leu	Tyr	Val	Pro 635	Pro	Gly	Ile	Phe	Lys 640	Ser	Thr	Cys	Lys	Met 645	Asp		
ATC	GCG	TGG	TTT	CCC	TTC	GAC	GAC	CAA	CAC	TGT	GAT	ATG	AAG	TTC	GGT	592	
Ile	Ala	Trp	Phe 650	Pro	Phe	Asp	Asp	Gln 655	His	Cys	Asp	Met	Lys 660	Phe	Gly		
AGC	TGG	ACA	TAT	GAC	GGC	AAT	CAG	TTG	GAT	CTG	GTG	CTA	AAA	GAT	GAG	640	
Ser	Trp	Thr 665	Tyr	Asp	Gly	Asn	Gln 670	Leu	Asp	Leu	Val	Leu 675	Lys	Asp	Glu		
GCA	GGC	GGC	GAT	CTA	TCG	GAC	TTC	ATA	ACA	AAT	GGG	GAG	TGG	TAT	CTA	688	
Ala	Gly 680	Gly	Asp	Leu	Ser	Asp 685	Phe	Ile	Thr	Asn	Gly 690	Glu	Trp	Tyr	Leu		
ATA	GGA	ATG	CCA	GGC	AAA	AAG	AAC	ACA	ATA	ACA	TAC	GCG	TGC	TGC	CCC	736	
Ile	Gly	Met	Pro	Gly	Lys 700	Lys	Asn	Thr	Ile	Thr 705	Tyr	Ala	Cys	Cys	Pro 710		
GAG	CCC	TAC	GTG	GAC	GTC	ACC	TTC	ACC	ATC	ATG	ATA	AGA	AGA	CGA	ACC	784	
Glu	Pro	Tyr	Val	Asp 715	Val	Thr	Phe	Thr	Ile 720	Met	Ile	Arg	Arg	Arg 725	Thr		
TTG	TAC	TAC	TTC	TTC	AAC	CTG	ATC	GTC	CCG	TGC	GTG	CTG	ATC	TCA	TCG	832	
Leu	Tyr	Tyr	Phe 730	Phe	Asn	Leu	Ile	Val 735	Pro	Cys	Val	Leu 740	Ile	Ser	Ser		
ATG	GCA	CTC	CTC	GGC	TTC	ACA	CTG	CCA	CCA	GAC	TCC	GGA	GAG	AAA	CTC	880	
Met	Ala	Leu 745	Leu	Gly	Phe	Thr	Leu 750	Pro	Pro	Asp	Ser	Gly 755	Glu	Lys	Leu		
ACA	CTT	GGA	GTC	ACT	ATT	CTT	CTA	TCG	CTG	ACG	GTG	TTC	CTC	AAC	CTG	928	
Thr	Leu 760	Gly	Val	Thr	Ile	Leu 765	Leu	Ser	Leu	Thr	Val 770	Phe	Leu	Asn	Leu		
GTA	GCC	GAG	ACC	CTG	CCA	CAG	GTC	TCC	GAC	GCT	ATC	CCC	CTG	TTA	GGG	976	
Val	Ala	Glu	Thr	Leu	Pro 780	Gln	Val	Ser	Asp	Ala 785	Ile	Pro	Leu	Leu	Gly 790		
ACG	TAC	TTC	AAT	TGC	ATC	ATG	TTC	ATG	GTA	GCG	TCG	TCT	GTG	GTA	CTG	1024	
Thr	Tyr	Phe	Asn 795	Cys	Ile	Met	Phe	Met	Val 800	Ala	Ser	Ser	Val	Val 805	Leu		
ACT	GTG	GTG	GTA	CTC	AAT	TAC	CAC	CAT	CGA	ACA	GCT	GAT	ATA	CAT	GAA	1072	
Thr	Val	Val	Val 810	Leu	Asn	Tyr	His	His 815	Arg	Thr	Ala	Asp	Ile 820	His	Glu		

ATG Met	CCA Pro	CAG Gln 825	TGG Trp	ATA Ile	AAA Lys	TCA Ser	GTA Val 830	TTC Phe	CTA Leu	CAA Gln	TGG Trp	TTG Leu 835	CCA Pro	TGG Trp	ATA Ile	1120
CTG Leu	CGA Arg 840	ATG Met	TCG Ser	AGG Arg	CCA Pro	GGG Gly 845	AAG Lys	AAG Lys	ATC Ile	ACC Thr	AGG Arg 850	AAG Lys	ACT Thr	ATA Ile	ATG Met	1168
ATG Met 855	AAC Asn	ACG Thr	AGG Arg	ATG Met	AGG Arg 860	GAG Glu	CTG Leu	GAA Glu	CTG Leu	AAG Lys 865	GAG Glu	AGG Arg	TCG Ser	TCG Ser	AAG Lys 870	1216
TCC Ser	TTG Leu	CTG Leu	GCG Ala	AAT Asn 875	GTT Val	CTA Leu	GAT Asp	ATT Ile	GAT Asp 880	GAT Asp	GAC Asp	TTC Phe	AGA Arg	CAC His 885	GGC Gly	1264
CCT Pro	CCG Pro	CCT Pro	CCT Pro 890	AAC Asn	AGT Ser	ACT Thr	GCC Ala	TCG Ser 895	ACC Thr	GGG Gly	AAT Asn	TTG Leu	GGA Gly 900	CCT Pro	GGG Gly	1312
TGC Cys	TCA Ser	ATA Ile 905	TTC Phe	CGC Arg	ACG Thr	GAT Asp	TTC Phe 910	CGT Arg	CGG Arg	TCG Ser	TTC Phe	GTC Val 915	CGT Arg	CCG Pro	TCC Ser	1360
ACG Thr	ATG Met 920	GAA Glu	GAC Asp	GTG Val	GGC Gly	GGC Gly 925	GGG Gly	CTG Leu	GGT Gly	AGC Ser	CAC His 930	CAT His	CGC Arg	GAG Glu	CTG Leu	1408
CAC His 935	CTC Leu	ATA Ile	CTG Leu	AGA Arg	GAG Glu 940	CTG Leu	CAG Gln	TTC Phe	ATC Ile	ACG Thr 945	GCC Ala	AGG Arg	ATG Met	AAG Lys	AAG Lys 950	1456
GCT Ala	GAT Asp	GAG Glu	GAA Glu	GCC Ala 955	GAG Glu	CTG Leu	ATC Ile	AGC Ser	GAC Asp 960	TGG Trp	AAG Lys	TTT Phe	GCT Ala	GCG Ala 965	ATG Met	1504
GTT Val	GTT Val	GAT Asp	AGG Arg 970	TTT Phe	TGC Cys	CTG Leu	TTC Phe	GTG Val 975	TTC Phe	ACA Thr	CTT Leu	TTC Phe	ACA Thr 980	ATC Ile	ATC Ile	1552
GCG Ala	ACA Thr	GTA Val 985	GCT Ala	GTC Val	CTG Leu	TTA Leu	TCG Ser 990	GCA Ala	CCG Pro	CAT His	ATC Ile	ATC Ile 995	GTG Val	CAA Gln		1597
TGAACCAACC ACTGAGCCGG CAACTCCGGC GCATGAATGA GAGAAATAAT TATTAGATCG																1657
CCGATTTGTA ATTATAATTG ATAATGTAAT TAAATTAAAT ACGTGTTGA AACGCACACG																1717
TCTCCATAAC AAAGTCTTAA GACATTAAAT TATGATAAAT TTACATATTG TAGTTAAGTC																1777
GAGTGTTGAT GGAAATTTTA GCCGGCGCAA GGAGTTTCGT GAAGGTCTGT ATATATTTTT																1837
TCTTATTGTT GTATATTGTA TCGTTGTTC																1897
GTTTGTCTCT TCGATGGCAG GTGCACTTCA GTTCAGGCTG AAATTTCCAT TAACATTTAT																1957
TTAAACAAAT GTGATGTTGA CTAGGATGTT ATACAGATAA ATGTTGACGT GTATAATTTG																2017
TTAAAATAAA CAATATTAAT TACTATTACT AAACGATATT ATAAACGAAG TACTAACGAG																2077
GGTTACTTTA ATGGGAAGAA CGCTAAGCTG GCACAGAGTT GCATTAATTT GAAAAAAGAA																2137
ATTACGGAAA AAAGTTTATT GAAAATTGAA CTTTTTGAA GGAAAGTAAC GTTTGATCAA																2197
AAAAGTTTGT AAAACGAAAG TTCGGTTCTG CGCCAATACT GGAATTAAAA TTCTCGTAAA																2257
TATTAGGGAA AAGAAGGTCC TTTAAAACAA AAGATTTGAA CCGGCATCCT TTTTACAAGT																2317

AATGAGGGAT CACAGATGAT GACAAAAAAC CTTAGGGTAT ATAAGTAATG TACATAATGG 2377
 ATCAAATATC GGTAGAGTCA AGAATAGTTA ACGATTTAAG ATTATTCCAT TCGATATTAA 2437
 AATTGATTA GCGATTGTCG CTGCGTCTAC TTTGATACAT ATCGATTTGA ATCGATATTG 2497
 TATAAATTTA GATAGATCGG ACATTAGTAA TGAGTATGGA CGTTTTAATT TTTAAAAAAG 2557
 AATGTACTAC GAAGATTAAA TCCAGGAATT GTTAAACAGT TATGGAATTG ATAAGAAATC 2617
 AACAATTAAT ACGGAACCAA AGGTAGACTA GGTGTAGCAT CAGGAGATTG AATTAAAACA 2677
 TAAATTAGGA CCGACTTAAA TGGAACCTGC GAGTGTATTG ATAACTTTTT AATTTAAAAA 2737
 CTCATTGTCG ATTAAATGGA GAATAACTTT TGATCTCTCG TATCGATAAA TGCTCACTTA 2797
 ACTATCGATA GCGTAATATT ATAACTGTTA GTATATCGAT ATGGGAGTAA GTCCTAGCA 2857
 TCAGAAATAG TCATTAATTA GGAATCGGTT TGTGTTAATG TTATGCTTAG CGAAAATATT 2917
 ACAATGCTGT TGATATCACT AACCATCACG TAACCATATT GATAAAATGT AAATACAGAA 2977
 TATTGCGGTG TGTATTGTA TATAAATTTT AGAAAAA AAAA AACTCGAGAG 3037
 TACTTCTAGA GCGGCCGCGG GCCCATCGAT TTTCCACCCG GGTGGGGTAC CAGGTAAGTG 3097
 TACCCAATTC GC 3109

(2) INFORMATION FOR SEQ ID NO: 6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 501 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) TYPE OF MOLECULE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

Met Ala Pro Met Leu Ala Ala Leu Ala Leu Leu Ala Leu Leu Pro Val
 1 5 10 15
 Ser Glu Gln Gly Pro His Glu Lys Arg Leu Leu Asn Ala Leu Leu Ala
 20 25 30
 Asn Tyr Asn Thr Leu Glu Arg Pro Val Ala Asn Glu Ser Glu Pro Leu
 35 40 45
 Glu Val Arg Phe Gly Leu Thr Leu Gln Gln Ile Ile Asp Val Asp Glu
 50 55 60
 Lys Asn Gln Leu Leu Ile Thr Asn Ile Trp Leu Ser Leu Glu Trp Asn
 65 70 75 80
 Asp Tyr Asn Leu Arg Trp Asn Asp Ser Glu Tyr Gly Gly Val Lys Asp
 85 90 95
 Leu Arg Ile Thr Pro Asn Lys Leu Trp Lys Pro Asp Val Leu Met Tyr
 100 105 110
 Asn Ser Ala Asp Glu Gly Phe Asp Gly Thr Tyr Gln Thr Asn Val Val
 115 120 125
 Val Arg Ser Gly Gly Ser Cys Leu Tyr Val Pro Pro Gly Ile Phe Lys
 130 135 140
 Ser Thr Cys Lys Met Asp Ile Ala Trp Phe Pro Phe Asp Asp Gln His

145		150		155		160
Cys	Asp	Met	Lys	Phe 165	Gly	Asp
Leu	Val	Leu	Lys 180	Asp	Glu	Ala
Asn	Gly	Glu 195	Trp	Tyr	Leu	Ile
Thr	Tyr 210	Ala	Cys	Cys	Pro	Glu
Met	Ile	Arg	Arg	Arg	Thr	Leu
Cys	Val	Leu	Ile	Ser 245	Ser	Met
Asp	Ser	Gly	Glu 260	Lys	Leu	Thr
Thr	Val	Phe 275	Leu	Asn	Leu	Val
Ala	Ile	Pro	Leu	Leu	Gly	Thr
Ala	Ser	Ser	Val	Val	Leu	Thr
Thr	Ala	Asp	Ile	His 325	Glu	Met
Gln	Trp	Leu	Pro 340	Trp	Ile	Leu
Thr	Arg	Lys 355	Thr	Ile	Met	Met
Lys	Glu 370	Arg	Ser	Ser	Lys	Ser
Asp	Asp	Phe	Arg	His	Gly 390	Pro
Gly	Asn	Leu	Gly	Pro	Gly	Cys
Ser	Phe	Val	Arg 420	Pro	Ser	Thr
Ser	His	His 435	Arg	Glu	Leu	His
Thr	Ala	Arg	Met	Lys	Lys	Ala
Trp	Lys	Phe	Ala	Ala	Met 470	Val
Thr	Leu	Phe	Thr	Ile 485	Ile	Ala
His	Ile	Ile	Val	Gln		

Patent Claims

1. Nucleic acid which comprises a sequence selected from
 - 5 (a) the sequences according to SEQ ID NO: 1, SEQ ID NO: 3 or SEQ ID NO: 5,
 - (b) part sequences, which are least 14 base pairs in length, of the sequences defined under (a),
 - 10 (c) sequences which hybridize with the sequences defined under (a) in 2 x SSC at 60°C, preferably in 0.5 x SSC at 60°C, particularly preferably in 0.2 x SSC at 60°C,
 - 15 (d) sequences which exhibit at least 70% identity with the sequences defined under (a), between position 1295 and position 2195 from SEQ ID NO: 1, or between position 432 and position 1318 from SEQ ID NO: 3, or between position 154 and position 1123 from SEQ ID NO: 5,
 - 20 (e) sequences which are complementary to the sequences defined under (a), and
 - (f) sequences which, on account of the degeneracy of the genetic code, encode the same amino acid sequences as the sequences defined under (a) to (d).
 - 25
2. Vector which comprises at least one nucleic acid according to Claim 1.
- 30 3. Vector according to Claim 2, characterized in that the nucleic acid molecule is functionally linked to regulatory sequences which ensure the expression of the nucleic acid in prokaryotic or eukaryotic cells.
4. Host cell which contains a nucleic acid according to Claim 1 or a vector according to Claim 2 or 3.
- 35

5. Host cell according to Claim 4, characterized in that it is a prokaryotic or eukaryotic cell.
- 5 6. Host cell according to Claim 5, characterized in that the prokaryotic cell is E.coli.
7. Host cell according to Claim 5, characterized in that the eukaryotic cell is a mammalian cell or an insect cell.
- 10 8. Polypeptide which is encoded by a nucleic acid according to Claim 1.
9. Acetylcholine receptor which comprises at least one polypeptide according to Claim 8.
- 15 10. Process for preparing a polypeptide according to Claim 8, which comprises
 - (a) culturing a host cell according to one of Claims 4 to 7 under conditions which ensure the expression of the nucleic acid according to Claim 1, and
 - 20 (b) isolating the polypeptide from the cell or the culture medium.
11. Antibody which reacts specifically with the polypeptide according to Claim 8 or the receptor according to Claim 9.
- 25 12. Transgenic invertebrate which contains a nucleic acid according to Claim 1.
13. Transgenic invertebrate according to Claim 12, characterized in that it is *Drosophila melanogaster* or *Caenorhabditis elegans*.
- 30 14. Process for producing a transgenic invertebrate according to Claim 12 or 13, which comprises introducing a nucleic acid according to Claim 1 or a vector according to Claim 2 or 3.
- 35 15. Transgenic progeny of an invertebrate according to Claim 12 or 13.

16. Process for preparing a nucleic acid according to Claim 1, which comprises the following steps:
- 5 (a) carrying out an entirely chemical synthesis in a manner known per se, or
- (b) chemically synthesizing oligonucleotides, labelling the oligonucleotides, hybridizing the oligonucleotides to the DNA of an insect cDNA library, selecting positive clones and isolating the hybridizing DNA from positive clones, or
- 10 (c) chemically synthesizing oligonucleotides and amplifying the target DNA by means of PCR.
- 15 17. Regulatory region which naturally controls transcription of a nucleic acid according to Claim 1 in insect cells and ensures specific expression.
18. Process for discovering novel active compounds for plant protection, in particular compounds which alter the conducting properties of receptors according to Claim 9, which comprises the following steps:
- 20 (a) providing a host cell according to one of Claims 4 to 7,
- (b) culturing the host cell in the presence of a compound or a sample which comprises a multiplicity of compounds, and
- 25 (c) detecting altered performance properties.
19. Process for discovering a compound which binds to receptors according to Claim 9, which encompasses the following steps:
- 30 (a) bringing a host cell according to one of Claims 4 to 7, a polypeptide according to Claim 8 or a receptor according to Claim 9 into contact with a compound or a mixture of compounds under conditions which permit interaction of the compound(s) with the host cell, the polypeptide or the receptor, and
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- (b) determining the compound(s) which bind(s) specifically to the receptors.

5 20. Process for discovering compounds which alter the expression of receptors according to Claim 9, which comprises the following steps:

10 (a) bringing a host cell according to one of Claims 4 to 7 or a transgenic invertebrate according to Claim 11 or 12 into contact with a compound or a mixture of compounds,

(b) determining the receptor concentration, and

15 (c) determining the compound(s) which specifically influence(s) the expression of the receptor.

20 21. Use of at least one nucleic acid according to Claim 1, one vector according to Claim 2 or 3, one regulatory region according to Claim 16 or one antibody according to Claim 11 for discovering novel active compounds for plant protection or for discovering genes which encode polypeptides which are involved in synthesizing functionally similar acetylcholine receptors in insects.

Figure 1

